



**Joint ICE, IEMA, CIWEM and National Telford Institute Event**

# **WETLAND SYSTEMS**

## **Storm Water Management Control**

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# Overview

- **Sustainable Flood Retention Basins**
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  - Methodology
  - Results and Discussion
  - Conclusions
- **Integrated Constructed Wetlands**
  - Background
  - Objectives
  - Materials and Methods
  - Results and Discussion
  - Conclusions

# Sustainable Flood Retention Basins

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- **Methodology**
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  - Self-organizing Map Model
- **Results and Discussion**
  - Most Important Variables
  - Prediction of SFRB Variables
  - Prediction of SFRB Types
- **Conclusions**

# Introduction

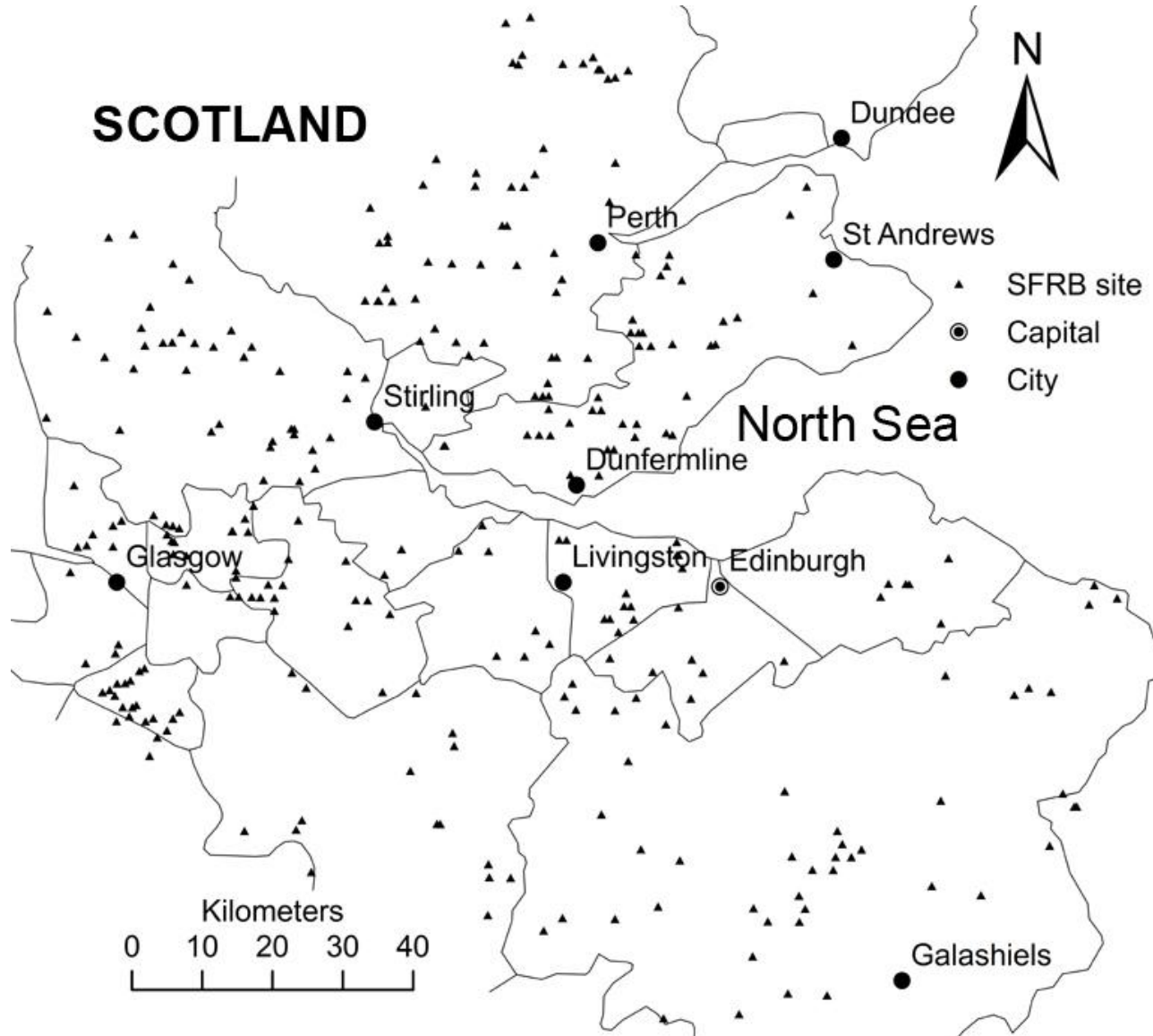
- The **EU Flood Directive 2007** requires member states to undertake **assessments** of artificial and natural **water bodies** including wetlands for their potential to contribute to **sustainable flood risk management** by 2013.
- A logical **concept** to **locate**, **classify** and **assess wetlands**, retention structures and flood defence structures would be highly valuable for **sustainable flood risk management planning**.

# Introduction

“A **Sustainable Flood Retention Basin** is defined as an impoundment or integrated **wetland**, which has a pre-defined or potential role in **flood and diffuse pollution control** that can be accomplished cost effectively through best management practice, **achieving sustainable flood risk management** and enhancing sustainable drainage, pollution reduction, biodiversity, green space, and recreational opportunities for society.” (Scholz, 2006)

# Introduction: Site Identification

Study area



# Introduction:

## Variables and Data Structure

### List of SFRB **variables**

ID	Variable and unit	ID	Variable and unit
1	Engineered (%)	21	Impermeable Soil Proportion (%)
2	Dam Height (m)	22	Seasonal Influence (%)
3	Dam Length (m)	23	Site Elevation (m)
4	Outlet Arrangement and Operation (%)	24	Vegetation Cover (%)
5	Aquatic Animal Passage (%)	25	Algal Cover in Summer (%)
6	Land Animal Passage (%)	26	Relative Total Pollution (%)
7	Floodplain Elevation (m)	27	Mean Sediment Depth (cm)
8	Basin and Channel Connectivity (m)	28	Organic Sediment Proportion (%)
9	Wetness (%)	29	Flotsam Cover (%)
10	Proportion of Flow within Channel (%)	30	Catchment Size (km <sup>2</sup> )
11	Mean Flooding Depth (m)	31	Urban Catchment Proportion (%)
12	Typical Wetness Duration (d a <sup>-1</sup> )	32	Arable Catchment Proportion (%)
13	Estimated Flood Duration (d a <sup>-1</sup> )	33	Pasture Catchment Proportion (%)
14	Basin Bed Gradient (%)	34	Forest Catchment Proportion (%)
15	Mean Basin Flood Velocity (cm s <sup>-1</sup> )	35	Natural Catchment Proportion (%)
16	Wetted Perimeter (m)	36	Groundwater Infiltration (%)
17	Maximum Flood Water Volume (m <sup>3</sup> )	37	Mean Depth of the Basin (m)
18	Flood Water Surface Area (m <sup>2</sup> )	38	Length of Basin (m)
19	Mean Annual Rainfall (mm)	39	Width of Basin (m)
20	Drainage (cm d <sup>-1</sup> )	40	Viniculture Catchment Proportion (%)

# Introduction: SFRB Types

- Hydraulic Flood Retention Basin (SFRB Type 1)
- Traditional Flood Retention Basin (SFRB Type 2)
- Sustainable Flood Retention Wetland (SFRB Type 3)
- Aesthetic Flood Treatment Wetland (SFRB Type 4)
- Integrated Flood Retention Wetland (SFRB Type 5)
- Natural Flood Retention Wetland (SFRB Type 6)

# Introduction: SFRB Types

- Drinking water and hydropower reservoirs (Type 1).
- Former drinking water reservoirs and traditional retention basins (Type 2).
- Sustainable drainage systems (Type 3).
- Constructed wetlands (Type 4).
- Artificial water bodies within parks (Type 5).
- Natural wetlands (Type 6).

# Introduction:

## Overall Aim

- To outline the SFRB concept and methodology for **using cost-effective** and reliable **variables** to **predict time-consuming, costly** and dependant **variables** with the aid of a **self-organizing map model**; and
- To outline the methodology on how to **predict the SFRB types** with the self-organizing map model.

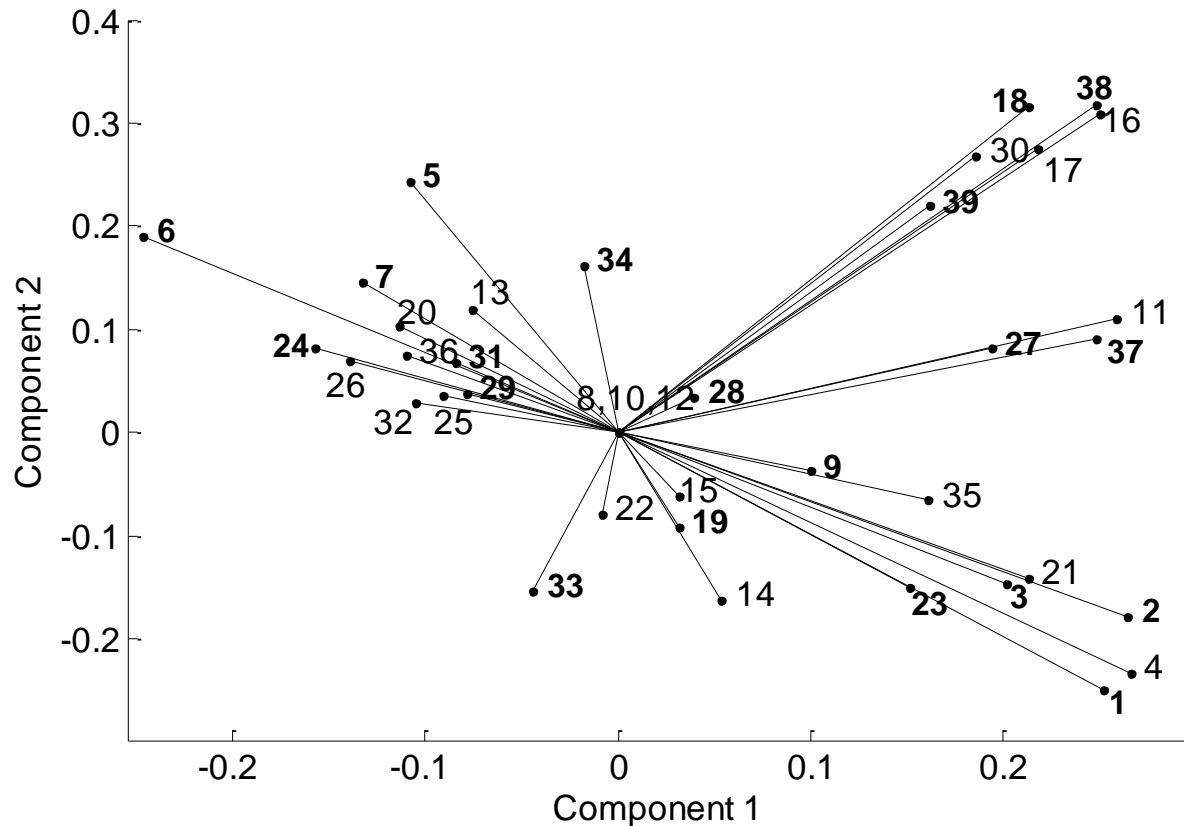
# Methodology

- Apply a **principal component analysis** to **identify relationships** between variables.
- Use the **self-organizing map** to **visualize** the **links** among variables.
- Apply the **self-organizing map** to **identify** the important and difficult to obtain variables and other easier-to-determine and inexpensive **variables**.

# Methodology

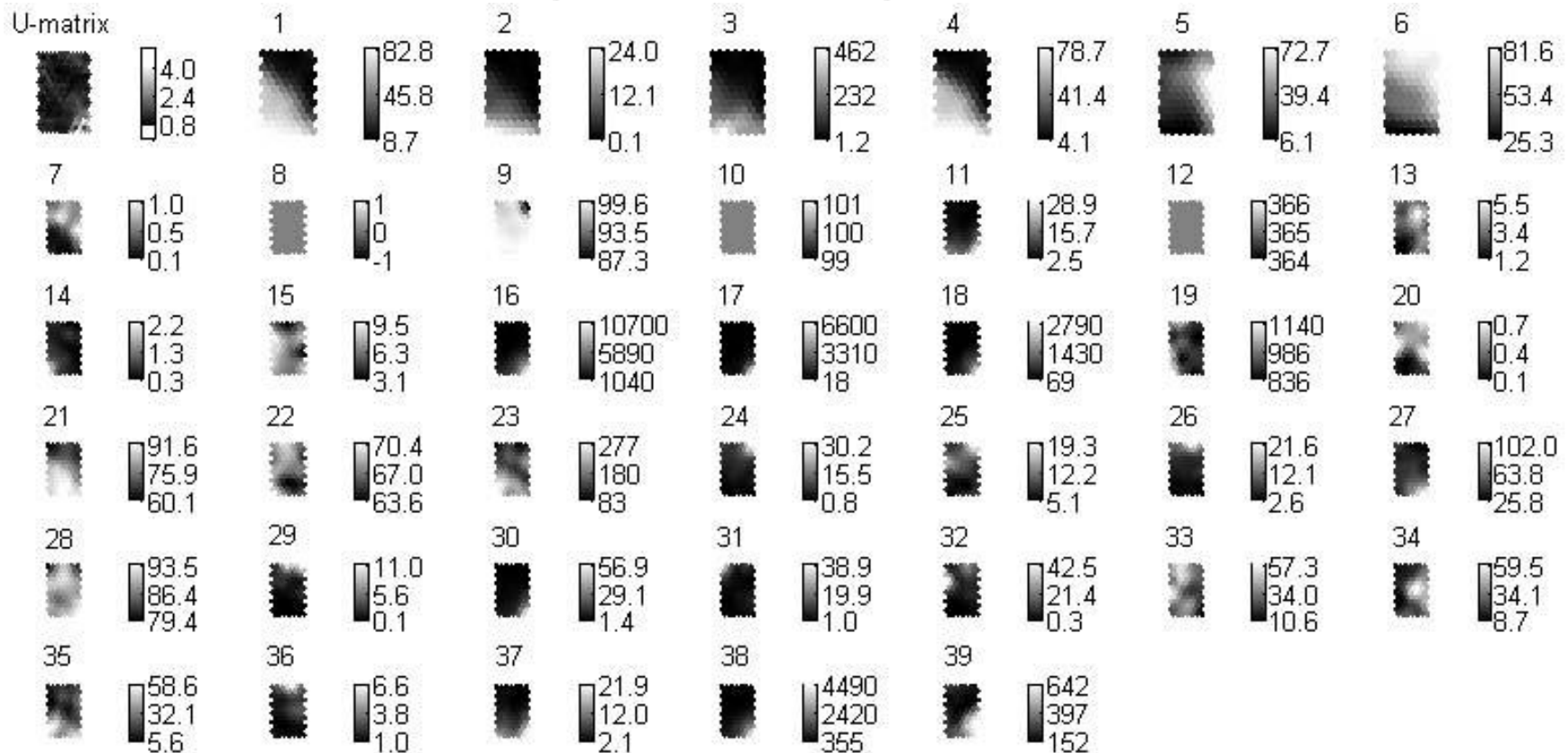
- Apply the self-organizing map model to **predict** difficult and expensive **variables** and **SFRB types**.
- Software package: **Matlab 7.1** for statistical analysis and modeling.

# Methodology: Principal Component Analysis



**Loading plot** of the **principal component analysis** identifying the most important classification variables and their similarities with each other

# Methodology: Self-organizing Map Model



**Self-organizing map:** Similar and corresponding patterns between variables indicate a close relationship; therefore, some variables can be eliminated from classification

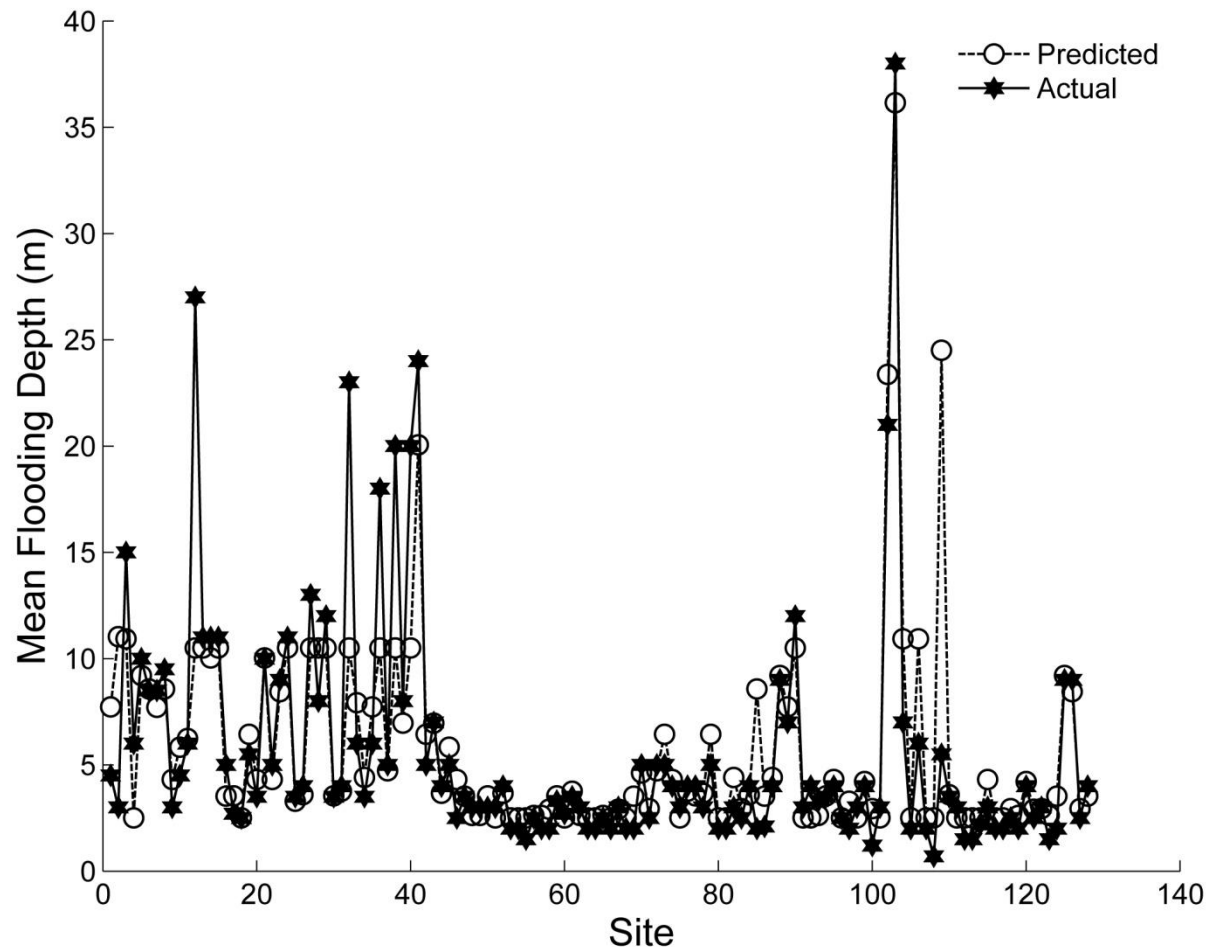
# Results and Discussion

- With the help of a **principal component analysis**, **20 groups of variables** (with similar contribution to classification) were **identified**.
- Therefore, **20 key variables** representing one group each were **identified**.
- The **self-organizing model** was successfully applied to **visualize** and subsequently **verify** the **relationships** identified by the principal component analysis.

# Results and Discussion

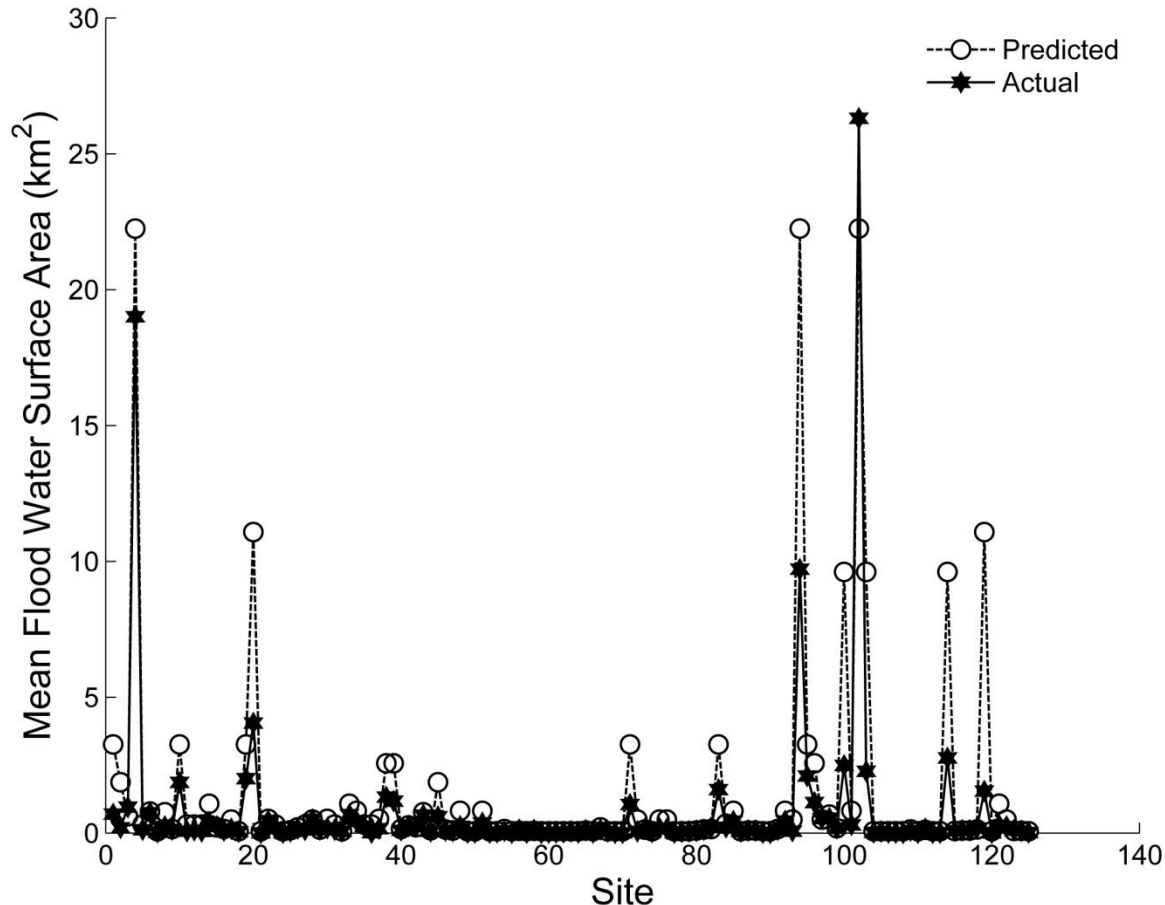
- For example, **low** data values for the variable ***Land Animal Passage*** are difficult to determine, but are **linked** to **high** values for the variables ***Engineered*** and ***Outlet Arrangement and Operation***.
- This finding **verifies** their **negative correlation** identified using principal component analysis.

# Results and Discussion



Using *Wetted Perimeter*, *Flood Water Surface Area*, *Catchment Size*, *Mean Depth of the Basin*, *Length of Basin*, and *Width of Basin* to predict **Mean Flooding Depth**

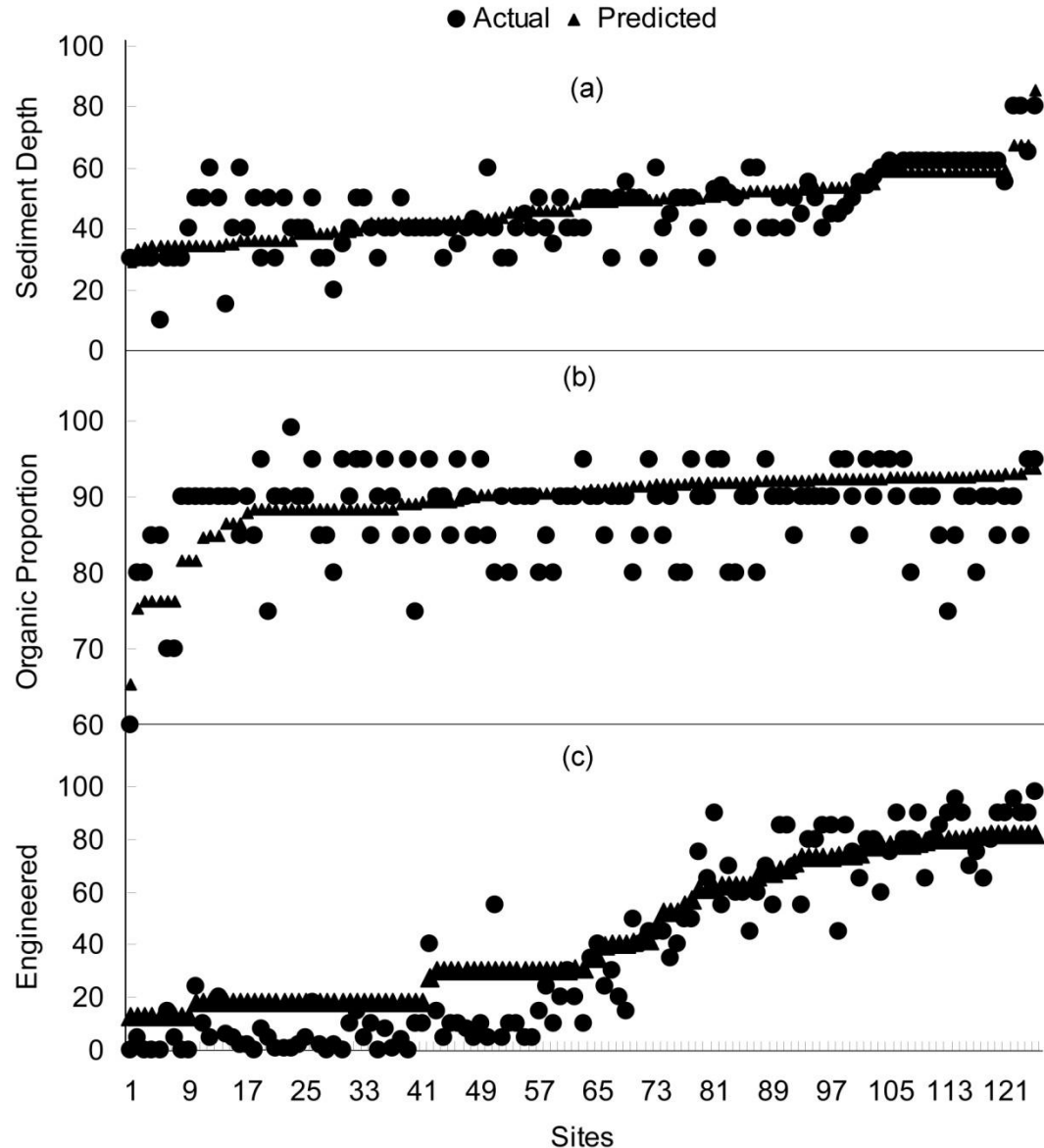
# Results and Discussion



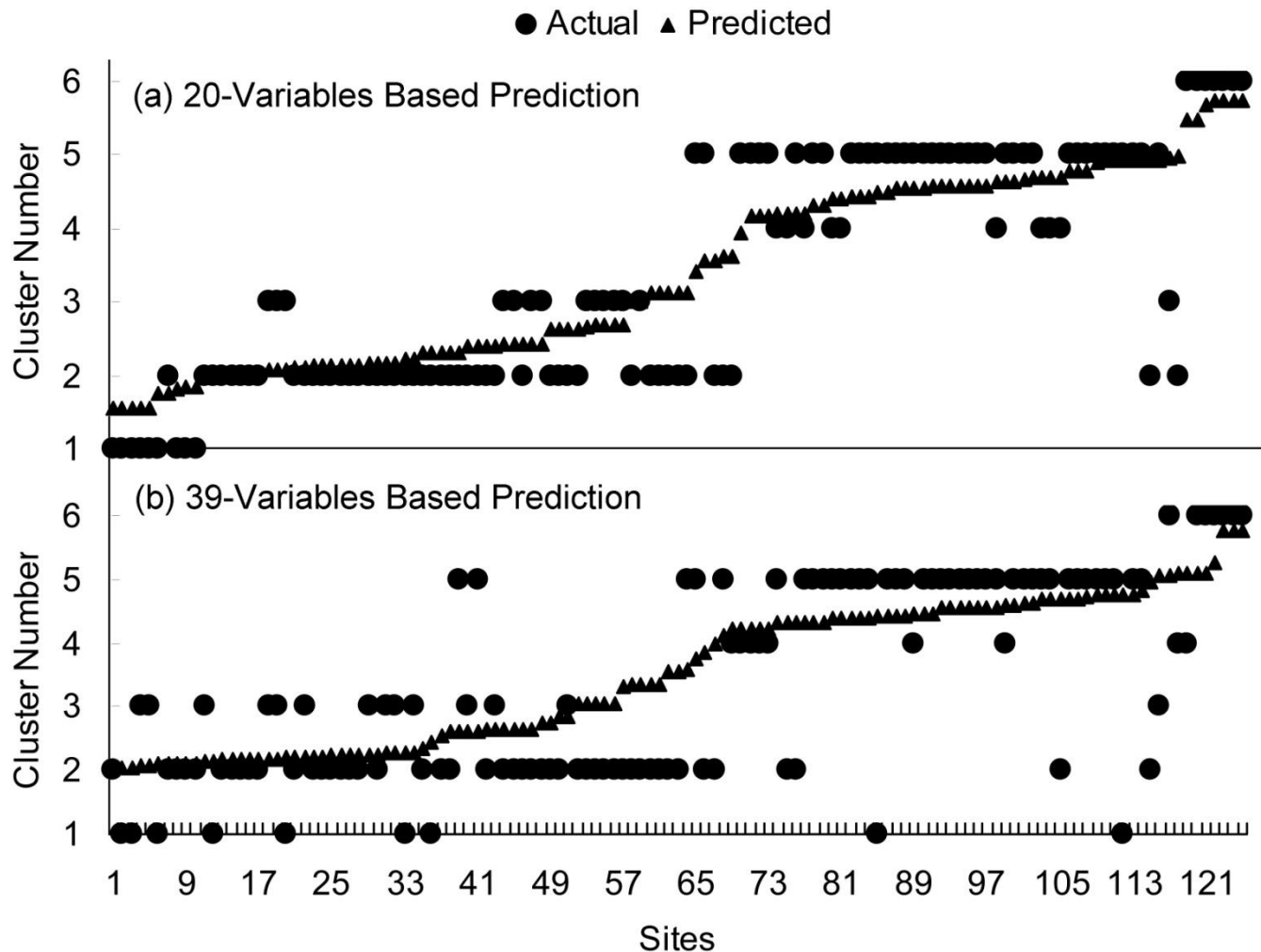
Using *Catchment Size, Mean Depth of the Basin, Length of Basin and Width of Basin* to estimate **Flood Water Surface Area**

# Results and Discussion

Prediction results for the variables (a) *Sediment Depth*, (b) *Organic Proportion* and (c) *Engineered*



# Results and Discussion



**Prediction of SFRB types** with the self-organizing map model based on (a) 20 and (b) 39 variables

# Conclusions

- The **20 most important variables** were identified to **classify water bodies** including wetlands and SFRB.
- The visualization of the relationships between variables displayed by the **self-organizing map** was **verified** the results of the **principal component analysis**.

# Conclusions

- Using highly correlated variables, which are easy to determine, to **predict costly** and difficult to obtain **variables** makes the SFRB assessment more **rapid** and **cost efficiently**.
- The self-organizing map model presents a **rapid and reliable** method of **predicting** different **types of water bodies** (i.e. SFRB types 1 to 6).

# Conclusions

- The **prediction model** can help to **avoid misunderstandings** concerning the water body status between different stakeholders and the scientific community.
- Learning the function and the status of water bodies will **help decision-makers** in **managing land use** in a sustainable manner and developing **flood risk management plans**.

# Integrated Constructed Wetlands

- Background
- Objectives
- Materials and Methods
- Results and Discussion
  - *Treatment Performance*
  - *Community Analysis*
  - *Bacterial Community Character*
  - *Microbial Community Assembly*
- Conclusions

# Background

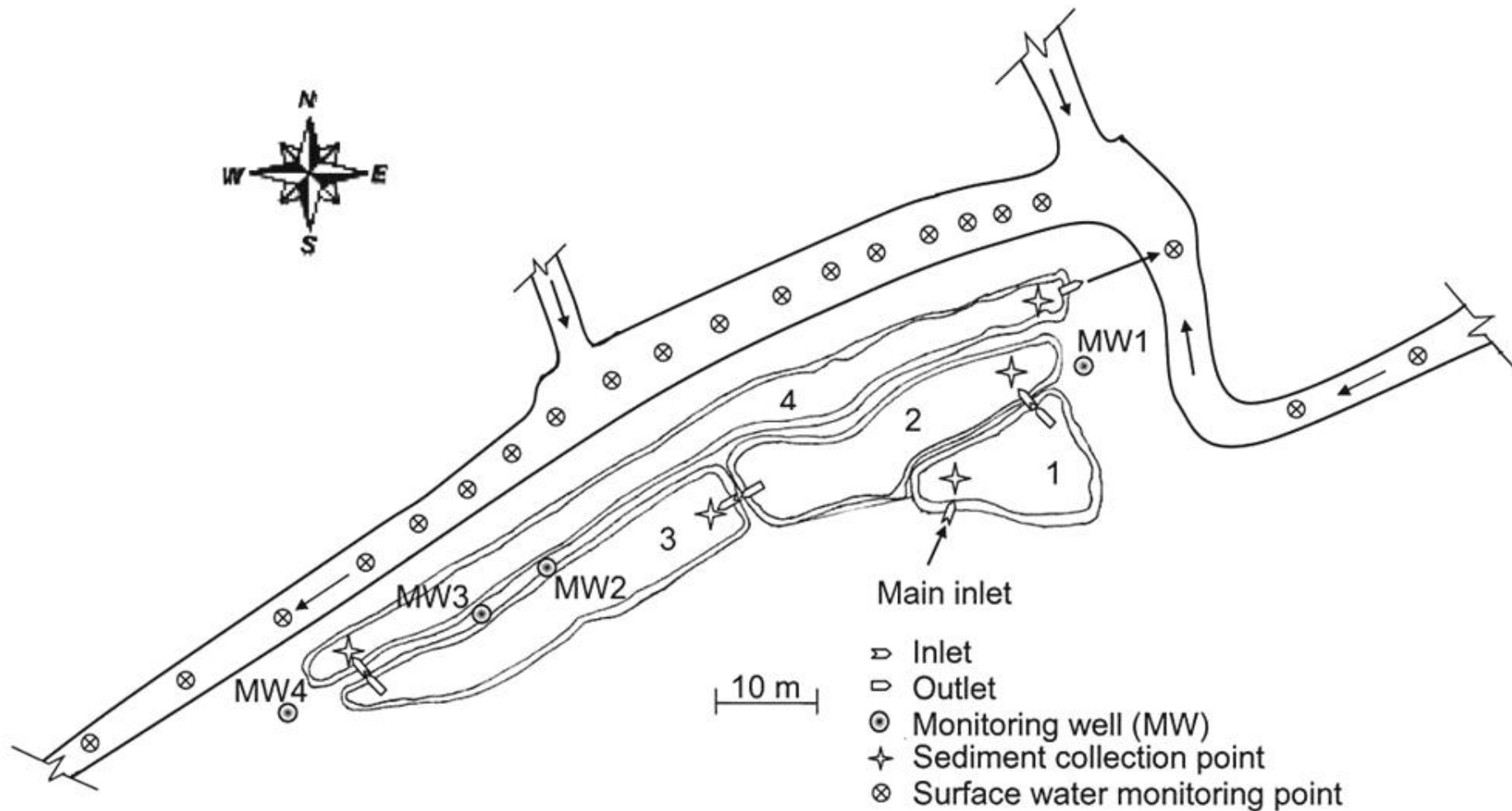
- **Non-point** sources of **water pollution** from anthropogenic sources such as those arising from intensive **farming** systems are a major environmental concern worldwide.
- Wastewaters from agricultural activities have high concentrations of **nutrients** and organic matter.
- **Constructed wetland** systems are low cost and an effective treatment technology to control the diffuse nature of agricultural wastewater pollution sources.

# Objectives

- To compare the **community composition** and **diversity of microorganisms** involved in nitrogen transformation from the **litter** and **sediments**.
- To assess a full-scale representative **ICW** system treating **ammonia-rich** wastewater.
- To **relate** differences in **diversity** to nitrogen **removal efficiencies**.
- To study differences in community composition have been analysed in the context of an ecological theory of **community assembly**.

# Materials and Methods

## *Study Site*



# Materials and Methods

## *Water Treatment*

- **Grab samples** for each wetland cell **inlet** and **outlet** were taken at an approximately **fortnightly** basis.
- Samples were analysed for pH, temperature, dissolved oxygen, biochemical oxygen demand, chemical oxygen demand, **ammonia-nitrogen**, **nitrate-nitrogen**, molybdate reactive phosphorus (soluble reactive phosphorus and *Escherichia coli*).

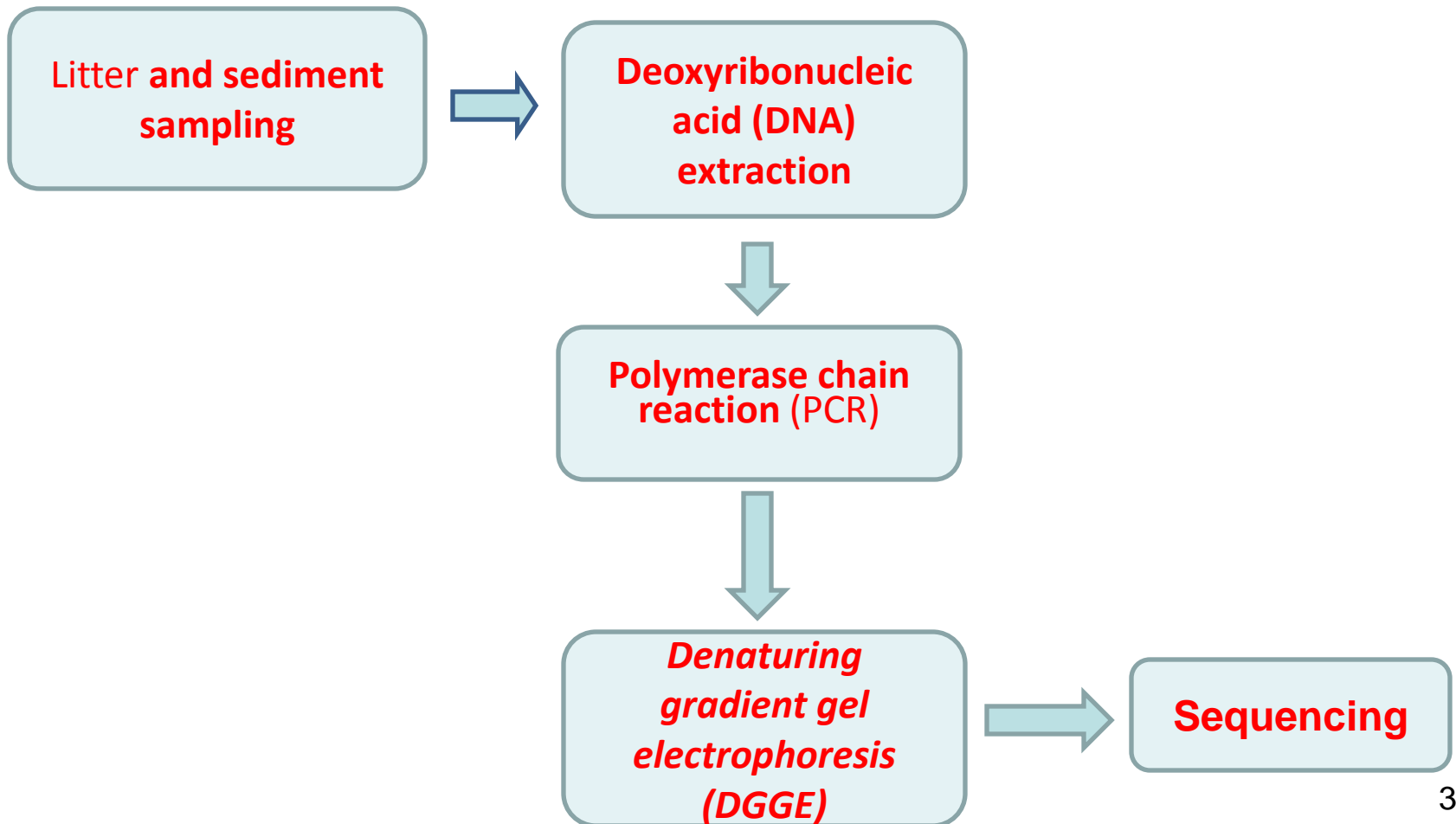
# Materials and Methods

## *Molecular Toolbox*

- Molecular methods were employed to study **ammonia-oxidisers** and putative **denitrifiers** in the wetland environment.
- **Polymerase chain reaction** (PCR) based methods were used for the nitrogen removing bacteria community analysis.

# Materials and Methods

## *Molecular Toolbox*



# Materials and Methods

## *Sample Collection*

- **Duplicate** field litter and sediment **samples** were collected from each wetland cell of the ICW system.
- For each sampling location, all **buried litter** in an area of **0.2 m<sup>2</sup>** was collected.
- Sediment samples were collected from the same area with a **sediment sampler** (**diameter of 4 cm**).
- The **upper 3 cm** of sediment located below the sediment-water interface were used for **analysis**.

# Materials and Methods

## *Sample Collection*

- The **samples** were collected near the **influent** point of each cell with an additional sample at the **outlet** of the last cell.
- All samples were **frozen immediately** after collection.
- Transported to the University of Newcastle for subsequent molecular microbiological analysis.

# Materials and Methods

## *DNA Extraction*

- The duplicate sediment and litter samples were subjected to deoxyribonucleic acid (**DNA**) **extraction**.
- Application of the **FastDNA® SPIN kit for Soil** (MP Biomedical Inc., USA) according to the manufacturer's protocol.

# Materials and Methods

## *PCR and Agarose Gel Electrophoresis*

- **Polymerase chain reaction** is a method to **multiply DNA segments** by repeating cycles of high and low temperature to separate DNA strands and to synthesize new strands.
- **Agarose gel electrophoresis** is a method to **separate DNA molecules by size**.

# Materials and Methods

## *PCR and Agarose Gel Electrophoresis*

- The **ammonia-oxidising bacterial** community was assessed using **primers** (Kowalchuk et al. 1997).
- The **denitrifying bacterial** community was assessed using **functional gene primers** (Throback et al., 2004).

# Materials and Methods

## *Denaturing Gradient Gel Electrophoresis (DGGE)*

- **DGGE** is a **molecular fingerprinting** method that **separates PCR-generated DNA** products.
- DGGE analyses were employed for the **separation of double-stranded DNA** fragments that are **identical in length**, but **differ in sequence**.
- **Polyacrylamide gels** (120×120×1 mm) were prepared with a denaturing gradient.

# Materials and Methods

## *Denaturing Gradient Gel Electrophoresis (DGGE)*

- The composition of **100% denaturant** was defined as 7M urea and 40% (vol/vol) formamide (Muyzer et al., 1993).
- The **gels were polymerised** with 15  $\mu\text{L}$  of TEMED and 150  $\mu\text{L}$  of ammonium persulphate.

# Materials and Methods

## *Sequencing*

- The **cleaned PCR products** were then **sequenced**.
- The **sequences** were then **BLAST analysed**.
- The internationally recognized NCBI BLAST ([www.ncbi.nih.gov](http://www.ncbi.nih.gov)) was used to **find closely related sequences** available in the public databases.

# Materials and Methods

## *Analysis of Denaturing Gradient Gel Electrophoresis*

- The **presence and intensity of bands** in the DGGE gels were analysed using **Bionumerics**.
- Comparisons of the **similarities** in community **composition** between different environmental sources (**leaves** or **sediments**) and **wetland cells** were conducted.
- Use of **non-metric multidimensional scaling** in combination with **cluster analysis** and **analysis of similarities**.
- Application of **Primer 6 for Windows**.

# Materials and Methods

- The **Bray-Curtis algorithm** was used to calculate pairwise **similarities** between samples using **presence-absence data**.
- The **Shannon-Weaver diversity index  $H$**  (applied on taxa abundance data; i.e. band intensity), **band richness  $S$**  and **Pielou's evenness  $J'$**  were calculated.
- These Parameters were used to **compare the diversity** of the predominant communities detected by DGGE.

# Materials and Methods

- **The nature of the assembly of the bacterial communities** within the ICW was **evaluated** using the Simberloff method, which is incorporated in to the **Raup and Crick similarity index** of the PAlaeontological Statistics (PAST) program.

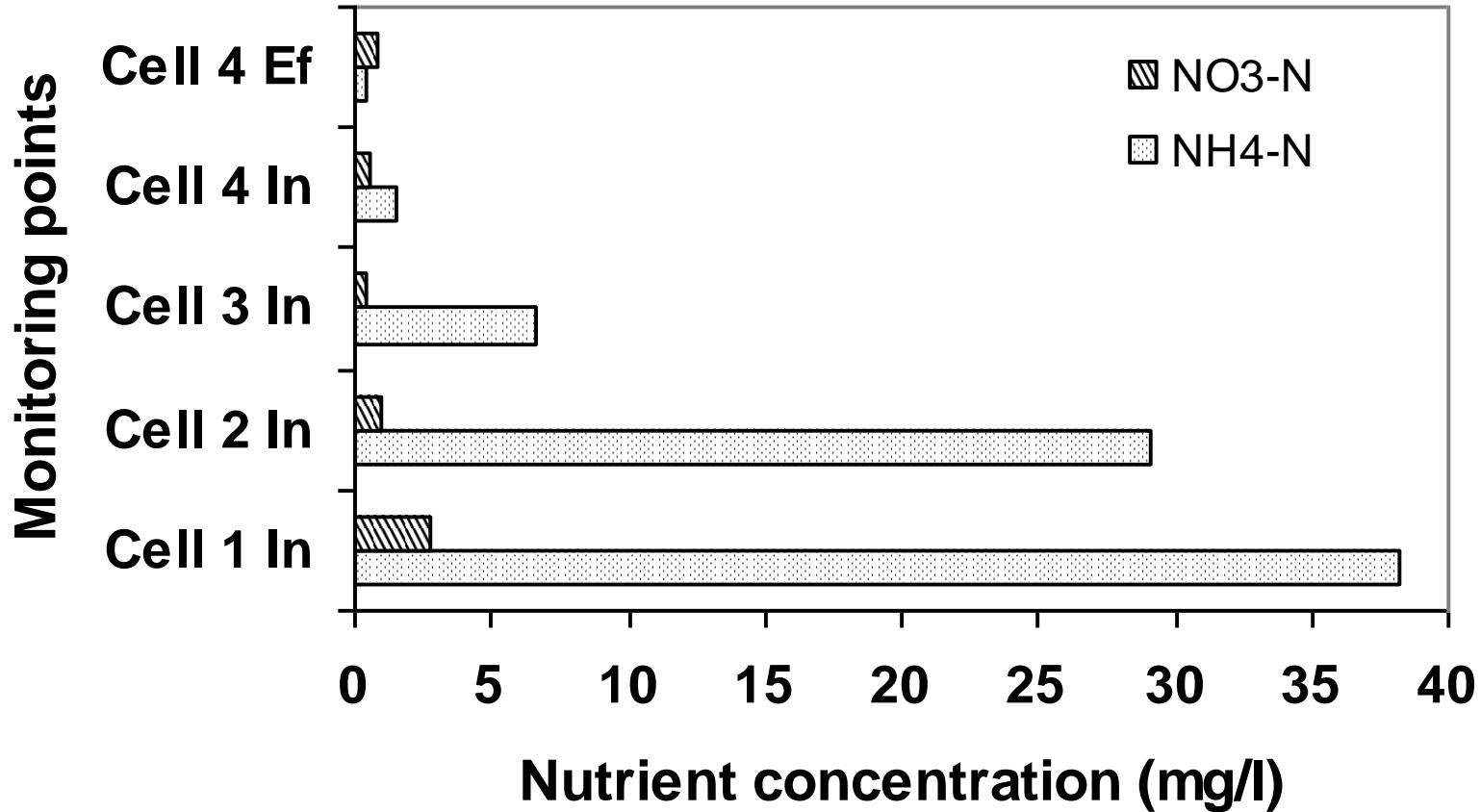
# Results and Discussion

## *Treatment Performance*

Parameter	ICW 11		
	Inlet	Outlet	RR %
Temperature (°C)	13.8	14.9	-
pH	8.12	7.37	-
Electrical Cond. (µS)	1469	373	-
SS (mg/l)	78.4	15.3	80.5
BOD <sub>5</sub> (mg/l)	593.1	5.8	99.0
COD (mg/l)	1341.5	50.4	96.2
<u>NH<sub>4</sub>-N (mg/l)</u>	<u>28.60</u>	<u>0.39</u>	<u>98.6</u>
<u>NO<sub>3</sub>-N (mg/l)</u>	<u>2.60</u>	<u>0.83</u>	<u>68.0</u>
MRP (mg/l)	8.13	0.83	89.8

# Results and Discussion

## *Nitrogen Removal Potential*

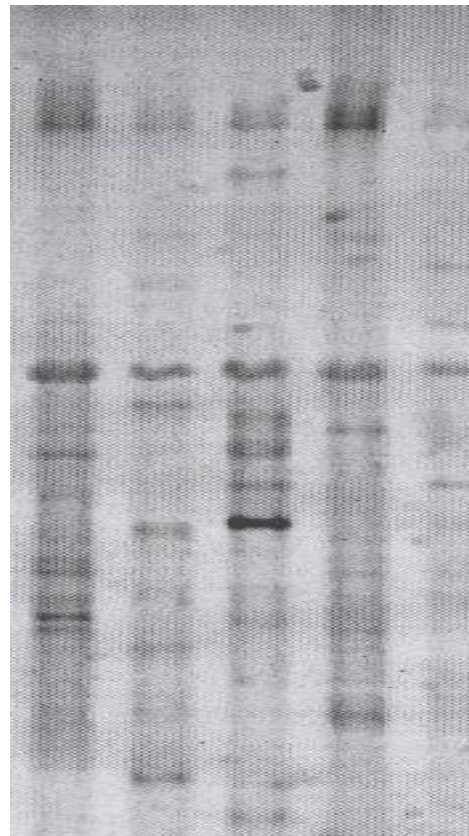


**Nutrient Reductions in Selected ICW Cells**  
(In, influent; Ef, effluent)

# Results and Discussion

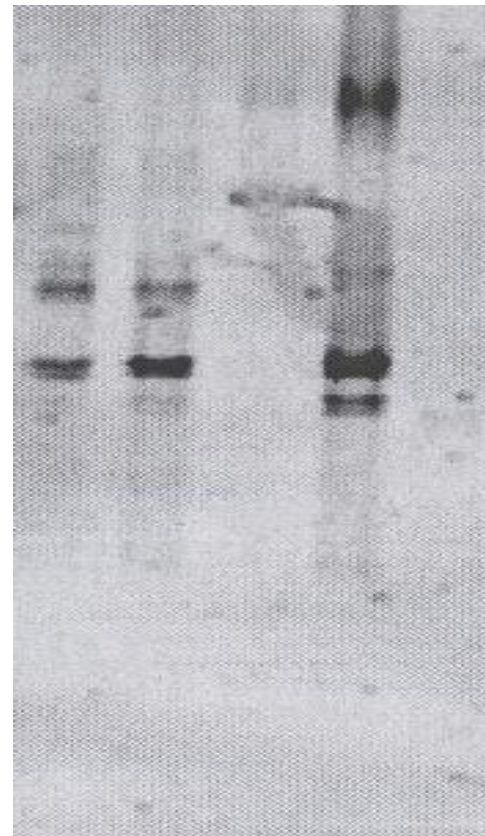
## *DGGE Profiles of PCR Products*

Denitrifying bacteria



C1 C2 C2 C3 C4

Ammonia oxidising bacteria



C1 C1 C2 C3 C4

# Results and Discussion

## *Community Analysis*

Sequence	ICW 11										Accession number	% Similarity	Strain
	C1 In		C2 In		C3 In		C4 In		C4 Ef				
	L	S	L	S	L	S	L	S	L	S			
C1			+		+						AY123811	97	Nitrosomonas sp. Nm59
C2	+	+			+			+			AY123801	99	Nitrospira sp. Nsp12
C3	+				+						AY727031	100	Nitrospira sp. En271
C4	+				+						AY792265	98	Uncultured beta proteobacterium clone

L= Litter

S= Sediment

# Results and Discussion

## Community Analysis

Sequence	ICW 11										Accession number	% Similarity	Strain	
	C1		C2		C3		C4		C4					
	In		In		In		In		Ef					
	L	S	L	S	L	S	L	S	L	S				
<i>nirK</i>														
C8	+		+									EU448024	81	Uncultured denitrifying bacterium clone T23_D5 nitrite reductase ( <i>nirK</i> ) gene
C9	+		+									EU448024	81	Uncultured denitrifying bacterium clone T23_D5 nitrite reductase ( <i>nirK</i> ) gene
C10	+		+									FM209186	86	<i>Pseudomonas aeruginosa</i> LESB58 complete genome sequence
C11			+									AY345247	78	<i>Pseudomonas aeruginosa</i> strain DN24 copper-dependent nitrite reductase
C12										+		EF623501	100	Uncultured bacterium clone LK22mK-28 nitrite reductase ( <i>nirK</i> ) gene
C13		+							+			AM230857	77	<i>Paracoccus</i> sp. R-26824 <i>nirK</i> gene for nitrite reductase
C14									+			DQ783326	96	Uncultured bacterium clone T1R2_0-7cm_038 <i>NirK</i> ( <i>nirK</i> ) gene
C16		+				+						AM419485	89	Uncultured organism partial <i>nirK</i> gene for putative copper containing dissimilatory nitrite reductase, clone Fin28
C18	+											EF615316	86	Uncultured bacterium clone P1m_ <i>nirK</i> -33 nitrite reductase ( <i>nirK</i> ) gene
C19	+											DQ337794	87	Uncultured bacterium clone S12m_ <i>nirK</i> -33 <i>NirK</i> ( <i>nirK</i> ) gene
C21	+		+		+					+		AM230832	82	<i>Rhizobium</i> sp. R-24663 <i>nirK</i> gene for nitrite reductase
C22			+									DQ337762	89	Uncultured bacterium clone P7m_ <i>nirK</i> -25 <i>NirK</i> -like ( <i>nirK</i> ) gene
C23	+		+		+							DQ304404	88	Uncultured bacterium clone Ag100-6 putative nitrite reductase ( <i>nirK</i> ) gene
<i>nirS</i>														
C24												AY078267	85	<i>Thauera terpenica</i> strain 21Mol putative dissimilatory nitrite reductase ( <i>nirS</i> ) gene,
C25	+		+		+			+				AM230919	90	<i>Dechloromonas</i> sp. R-28451 <i>nirS</i> gene for nitrite reductase
C26	+		+									AM230913	84	<i>Dechloromonas</i> sp. R-28400 <i>nirS</i> gene for nitrite reductase

# Results and Discussion

## *Diversity*

**Diversity indices** for the ammonia-oxidising and denitrifying bacterial communities in sediment and litter of the ICW system (mean  $\pm$  SD)

<b>Primer/ Genes</b>	<b>Component</b>	<b>Shannon's Index (H)</b>
<b>CTO (Ammonia- oxidisers)</b>	<b>Litter</b>	<b>0.68 <math>\pm</math> 0.80</b>
	<b>Sediment</b>	<b>n.d.</b>
<b><i>nirK</i> (Denitrifiers)</b>	<b>Litter</b>	<b>2.04 <math>\pm</math> 0.29</b>
	<b>Sediment</b>	<b>0.89 <math>\pm</math> 0.80</b>
<b><i>nirS</i> (Denitrifiers)</b>	<b>Litter</b>	<b>2.31 <math>\pm</math> 0.18</b>
	<b>Sediment</b>	<b>1.60 <math>\pm</math> 0.68</b>

n.d. no data

# Results and Discussion

## *Bacterial Community Characterisation*

- Putative **ammonia-oxidising bacteria** were **detected** by PCR in **all wetland cells except** for the **last cell** of the ICW, and at **more locations** in **leaf litter than** in **sediments**.
- **Denitrifiers** were **detected** in **all samples**.
- Analysis of putative **ammonia-oxidising bacteria** and **denitrifier** community fingerprints showed that there were **significant** (ANOSIM;  $p < 0.05$ ) **dissimilarities between communities**.

# Results and Discussion

## *Microbial Community Assembly*

- A comparison of microbial communities between the wetland cells suggested that the **ammonia-oxidising bacteria** and **denitrifying bacterial populations** were **no more similar or dissimilar than if they had been assembled by chance.**
- Most of the **Raup and Crick indices** were between **0.05 and 0.95.**

# Conclusions

- For **ammonia-oxidizing bacteria**, both *Nitrospira* and *Nitrosomonas* were detected in the studied wetland system.
- Concerning **denitrifying bacteria**, *Paracoccus*, *Pseudomonas*, *Rhizobium* and *Dechloromonas* were identified.
- The **litter** component of the studied wetland system **supported more diverse nitrogen removing bacteria** (ammonia-oxidising and denitrifying) **than** the **sediments**.

# Conclusions

- **Nitrogen removing bacteria** communities in the full-scale wetland system were **no more similar or dissimilar than** would be expected **by chance alone**.
- This suggests that stochastic assembly processes may be important in shaping these communities.

# Acknowledgements

- The **European Regional Development Fund Interreg IVB 2007-2013** funded this research project.
- The **Chinese Scholarship Council** and The University of Edinburgh Joint Scholarship supports Ms. Qinli Yang's PhD study.
- **University of Munich**, Germany.
- **Co-workers**: Chris Sagar, Louise Blackhall, Mark Keane, Rodney Moody and Michelle Robinson.

# Acknowledgements

- **Paul Carroll** and **Susan Cook**, Waterford County Council, Ireland.
- **Andy Gray** and **John Norman**, University of Edinburgh.
- Department of Environment, Heritage and Local Government, Ireland.
- The University of Edinburgh Development Trust.

# Further Reading

Miklas Scholz

GREEN ENERGY AND TECHNOLOGY

## Wetland Systems

Storm Water  
Management Control

 Springer

Thank you for your attention!